

### Introduction

- Rising atmospheric CO<sub>2</sub> concentration may lead to an increased input of available C from plants to the soil through rhizodeposition and may affect soil microbes with implications for the interaction between the C- and N-cycling.
- Changes of soil functional microbes associated with C- and N-cycling under long-term elevated CO<sub>2</sub> level (eCO<sub>2</sub>) suggest concomitant alterations of microbial biomass, N availability and gaseous N emissions.

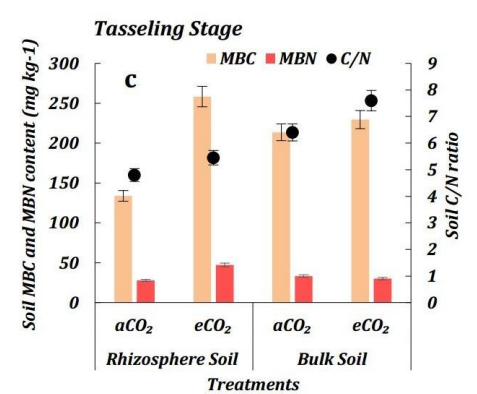


Fig.1 Effects of eCO<sub>2</sub> on MBC and MBN

### Objectives

- To investigate the impacts of 10-year eCO<sub>2</sub> on the microbial abundance and composition in both rhizospheric and bulk soils, based on functional marker genes for ammonia oxidation (bacterial *amoA*) and denitrification (*nirK*, *nirS*, *nosZ*), as well as fungi (ITS) and bacteria (16S rRNA).
- To examine the responses of soil microbial biomass (microbial biomass-C and -N) and soil N availability (mineral N) in both rhizospheric and bulk soils to long-term eCO<sub>2</sub>.

Table 1. The copy number of 16s rDNA and ITS in rhizospheric and bulk soil of maize at tasseling stage under elevated CO<sub>2</sub> concentration. Data are the mean ± S.E., n = 3. Different letters indicate significant differences among CO<sub>2</sub> concentration, rhizosphere and bulk soil at p < 0.05.

Treatment	16s rDNA (*10 <sup>9</sup> )	ITS (*10 <sup>7</sup> )	16s rDNA/ITS
aCO <sub>2</sub> -R	2.36 ± 0.28 a	1.58 ± 0.05 b	149.38 b
eCO <sub>2</sub> -R	1.77 ± 0.10 b	2.01 ± 0.16 a	88.67 c
aCO <sub>2</sub> -B	1.37 ± 0.30 c	0.16 ± 0.02 d	547.90 a
eCO <sub>2</sub> -B	1.95 ± 0.26 b	1.21 ± 0.09 c	137.35 b

Table 2. The copy number of AOB, nirS, nirK and nosZ in rhizospheric and bulk soil of maize at tasseling stage under elevated CO<sub>2</sub> concentration. Data are the mean ± S.E., n = 3. Different letters indicate significant differences among CO<sub>2</sub> concentration, rhizosphere and bulk soil at p < 0.05.

Treatment	AOB (10 <sup>7</sup> )	nirS (*10 <sup>6</sup> )	nirK (*10 <sup>9</sup> )	nosZ (*10 <sup>8</sup> )	nosZ/ (nirS + nirK)
aCO <sub>2</sub> -R	6.38 ± 0.10 b	2.77 ± 0.17 d	4.64 ± 0.30 b	4.75 ± 0.48 c	0.16 b
eCO <sub>2</sub> -R	6.56 ± 0.39 b	6.31 ± 0.37 b	7.63 ± 0.13 a	25.41 ± 5.53 b	0.59 a
aCO <sub>2</sub> -B	6.58 ± 0.99 b	3.54 ± 0.39 c	5.29 ± 0.82 ab	8.56 ± 0.84 c	0.10 b
eCO <sub>2</sub> -B	7.54 ± 0.23 a	9.29 ± 0.54 a	6.39 ± 0.55 ab	52.3 ± 2.51 a	0.33 ab

### Materials & Methods

#### 1. FACE experiment

- Ambient CO<sub>2</sub> (aCO<sub>2</sub>, 400 ppm)
- Elevated CO<sub>2</sub> (eCO<sub>2</sub>, 550 ppm)

#### 2. Studied soil

- Rhizospheric soil and bulk soil at 0-20 cm depth at tasseling stage of maize growth period

#### 3. Measurements

- Ammonium and nitrate concentrations
- Microbial biomass-C and -N (MBC and MBN)
- Quantity of microbial groups based on qPCR
- Composition of microbial communities based on DNA sequencing.

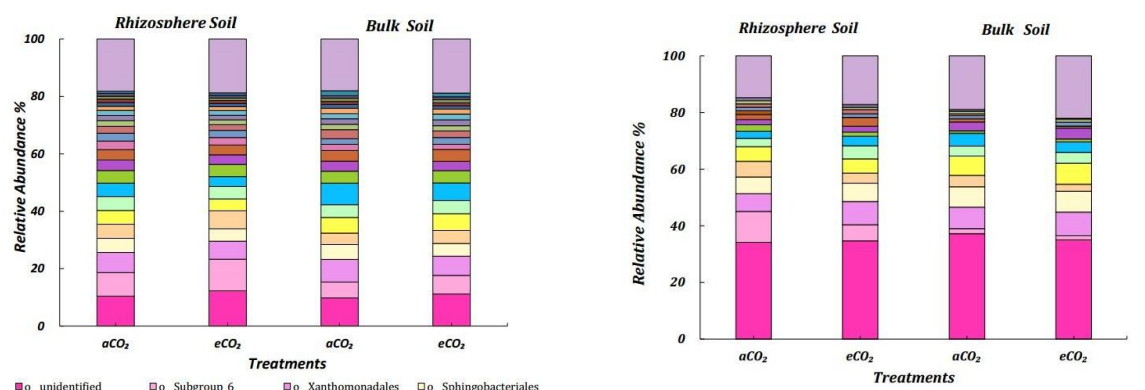


Fig.2 The composition of 16S rRNA at order level

Fig.3 The composition of ITS at genus level

### Results & Discussion

- MBC and MBN in rhizospheric soil were significantly increased under eCO<sub>2</sub>. This may be due to the increased roots exudates and root exfoliations induced by eCO<sub>2</sub>.
- In rhizospheric soil, increased quantity of fungi under eCO<sub>2</sub> and reduced bacterial quantity were observed, illustrating that the effect of eCO<sub>2</sub> mainly reflected in the increase of fungi.
- For bacterial community, *Sphingomonadales* that can produce catalase, was declined in relative abundance under eCO<sub>2</sub>, suggesting that oxygen content may be altered under eCO<sub>2</sub>. For fungal communities, *Chaetomium* and *Humicola* that can synthesize cellulase, hemicellulase and amylase, were increased in relative abundance under eCO<sub>2</sub>, possibly due to the increase in dead root litter.
- The quantity of AOB and denitrifiers were promoted under eCO<sub>2</sub>, particularly in rhizospheric soil.
- The changes of microbial community compositions associated with N-cycling were reflected on the decrease in *Nitrosospora* for AOB, increase in *Mesorhizobium* for *nirK*, increase in *Herbaspirillum* and *Bradyrhizobium* for *nosZ*.

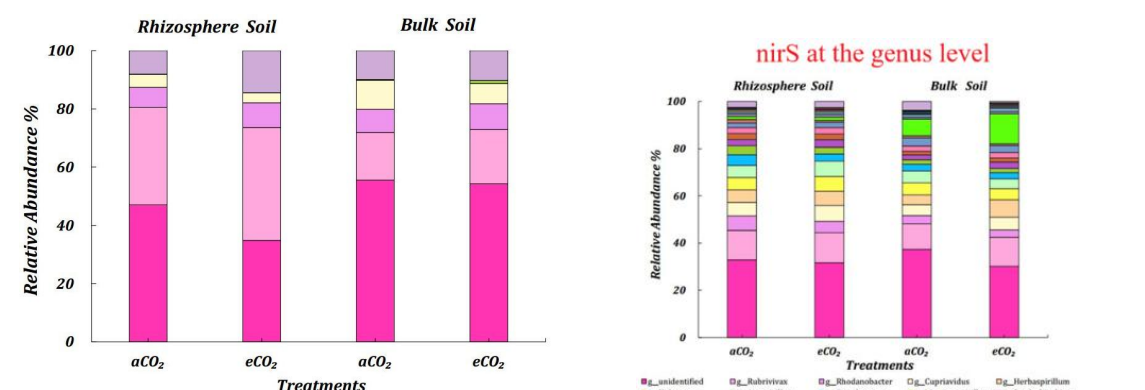
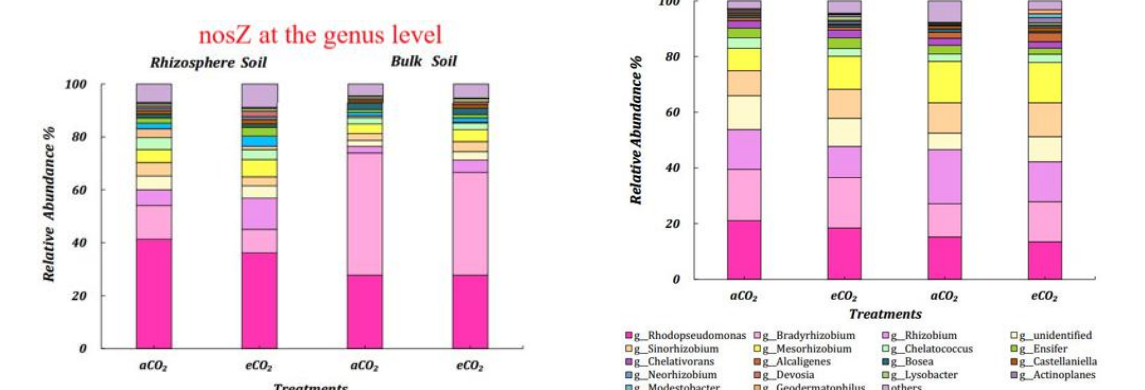


Fig.4 The composition of AOB at genus level

Fig.5 The composition of nirS, nirK and nosZ



### Conclusion

Ten years of CO<sub>2</sub> enrichment did not significantly change the community compositions of functional microbes associated with C- and N-cycling, possibly due to the differences in the form and quantity of soil C and N under eCO<sub>2</sub>, especially in rhizospheric soil.

### References

- Barnard, R., Barthes, L., Le Roux, X., Harmens, H., Raschi, A., Soussana, J.F., Winkler, B., Leadley, P.W., 2004. Atmospheric CO<sub>2</sub> elevation has little effect on nitrifying and denitrifying enzyme activity in four European grasslands. *Glob. Change Biol.* 10, 488-497.
- Carney, K.M., Hungate, B.A., Drake, B.G., Megonigal, J.P., 2007. Altered soil microbial community at elevated CO<sub>2</sub> leads to loss of soil carbon. *Proc. Natl. Acad. Sci.* 104, 4990-4995.
- Drigo, B., Kowalchuk, G.A., van Veen, J.A., 2008. Climate change goes underground: effects of elevated atmospheric CO<sub>2</sub> on microbial community structure and activities in the rhizosphere. *Biological Fertility Soils.* 44, 667-679.
- Drissner, D., Blum, H., Tscherko, D., Kandeler, E., 2007. Nine years of enriched CO<sub>2</sub> changes the function and structural diversity of soil microorganisms in a Grassland. *Eur. J. Soil Sci.* 58, 732-740.
- Phillips, R.P., Finzi, A.C., Bernhardt, E.S., 2011. Enhanced root exudation induces microbial feedbacks to N cycling in a pine forest under long-term CO<sub>2</sub> fumigation. *Ecol. Lett.* 14, 187-194.
- Zak, D.R., Pregitzer, K.S., Curtis, P.S., Holmes, W.E., 2000. Atmospheric CO<sub>2</sub> and the composition and function of soil microbial communities. *Ecological Applications.* 10, 47-59.